

Angiogenesis in Inflammatory Bowel Disease

Antoni Stadnicki^{1,2,*}, Anna Stadnicka^{3,4} and Wioletta Pollok-Waksmanska⁴

¹Faculty of Medicine, Collegium Medicum, Jan Dlugosz University in Czestochowa, Poland

²Polonia University in Czestochowa, Czestochowa, Poland

³District Hospital, Kolobrzeg, Poland

⁴Postgraduate student, CBT EDU Center, School of Cognitive-Behavioral Therapy in Szczecin, Warshaw, Poland

⁵Faculty of Health Sciences, University of Bielsko-Biala, Bielsko-Biala, Poland

***Corresponding author:**

Antoni Stadnicki,
Faculty of Medicine, Collegium Medicum, Jan Dlugosz
University in Czestochowa, Poland and Polonia Uni-
versity in Czestochowa, Czestochowa, Poland

Received: 03 Jan 2026

Accepted: 13 Jan 2026

Published: 20 Jan 2026

J Short Name: WJGHE

Copyright:

©2026 Antoni Stadnicki. This is an open
access article distributed under the terms of the
Creative Commons Attribution License, which permits
unrestricted use, distribution, and build upon your work
non-commercially

Keywords:

Inflammatory Bowel Disease; IBD; Angiogenesis; VEGF; Intestinal Barrier; Microbiota; Coagulation; Bradykinin

Citation:

Antoni Stadnicki, Angiogenesis in Inflammatory Bowel Disease. World Jour of Gastro and Hepatology® 2026; V10(5): 1-10

1. Abstract

The ethiology of inflammatory bowel diseases (IBD) is not precisely defined; however, they display environmental factors, genetic predisposition, gut microbiota involvement, and abnormal immunity. Angiogenesis seems to be an integral part of IBD. Impaired intestinal barrier probably expresses an initiating or early feature in the disease. A disruption of epithelial barrier leads to penetration of microbiota and other antigens into the mucosa leading to an enhanced immune response, whereas the vascular barrier damage is related to endothelium activation and pathological angiogenesis which promote inflammation. Angiogenesis is a very complex phenomenon which including endothelial and immune cells, growth factors, cytokines, adhesion molecules, intestinal microbiota, and signal transduction. It seems that intestinal microvasculature hemostasis turns to prothrombotic state, and microthrombi formation enhance ischemia. The angiogenic process in IBD is in part regulated by intestinal microbiota. Antiangiogenic therapy is a novel and significant approach for IBD treatment. Biologic anti-inflammatory therapy of IBD simultaneously attenuates angiogenesis to a similar degree. However expression of VEGF and other growth factors may have dual and opposing effect probably related to stage of the disease. Thus anti-angiogenic treatment in IBD patients is still controversial and clinical trials should be performed using anti-angiogenic drugs.

2. Introduction

Crohn's disease and ulcerative colitis (UC) are two entities of inflammatory bowel disease (IBD) with chronic local and systemic inflammatory changes and spontaneous relapsing course. The etiology and pathogenesis of IBD are still not fully known, but they show genetic and environmental factors and display immunologic alterations caused in part by intestinal microbiome [1]. IBD is clas-

sically considered as a disease of Western countries, but over the past few decades the incidence of the disease rapidly increased in newly industrialized Word regions. UC primarily impacts the large bowel, whereas Crohn's disease may affect any part of the gastrointestinal tract, from the oral cavity to the rectum. About 20-30% of IBD cases are associated with extraintestinal complications [1]. Intestinal barrier is created by epithelial and vascular layers to potentially harmful components. When the intestinal barrier is disrupted, bacterial and other antigens may enter the usually sterile submucosa and change immunological response, thus may play a role in the pathogenesis of IBD. The normal immunologic reactions suppress inflammation, however in genetically susceptible subjects inflammatory response is altered and amplified with defective role and regulation of innate immune cells, faulty epithelial barrier function and pathological angiogenesis. Extensive tissue injury is characterized by expression of adhesive molecules in the vessel wall with inflammatory cell infiltration, release of cytokines and other inflammatory mediators. It also may perpetuate of the commensal microbiota composition contribute to dysbiosis [2]. In the last years many reports have presented a role of intestinal microbiota in intestinal homeostasis and intestinal barrier dysfunction in IBD. In light of the recent evidence, this review is mainly focused on a proangiogenic activators, contribution of microbiota to activation of intestinal microvasculature, relation between dysregulation of intestinal barrier and pathological angiogenesis, and summarizes the effectiveness of anti-angiogenic therapy in IBD.

3. Angiogenesis

Angiogenesis, the process of new vessel formation, physiologically takes place during embryonic development and menstrual cycle. However in pathology formation of new vessels may persists which can induce or augment several pathological conditions as

adhesion molecule (JAM)-A, ocludin, and cingulin [31]. In addition, claudin family molecules; claudin-3, -5, and -12 and enteric glial cells which are expressed in intestinal endothelia [40, 41] develop the gut-vascular barrier which regulates soluble paracellular transport, compared to the brain-blood barrier (BBB) [42, 43]. However in contrast to BBB, which has a size exclusion threshold of 500 Da, the gut – vascular barrier is permeable to molecules as large as 4 kDa [31]. Thus intestinal permeability is mediated by both the epithelial and endothelial barriers disturbances.

5. Intestinal Hemostasis in IBD

An ischemic inflammation in the intestinal microvasculature due to pro-coagulant activity and thrombi formation further enhance tissue damage and pathological angiogenesis. Long time ago Wakefield et al [44, 45] observed granulomatous vasculitis in intestinal microvasculature during early stage of Crohn's disease, which suggests its pathogenic significance. Capillary thrombi linked with fibrin, and an expression of the tissue factor were also observed in intestinal vasculature in Crohn's disease and UC [46, 47]. Activated platelets shown in intestinal microcirculation of IBD patients may activate endothelial cells via the expression of CD40. Danese et al [48] have shown higher platelet expression of surface CD40 ligand (CD40L), and an elevated concentration of platelets origin soluble CD40L in plasma of UC and Cronh's disease patients as compared to healthy subjects. They also observed CD40L positive platelets adherents to ECs in intestinal circulation, which may initiate inflammatory response [49]. In addition, the interaction of platelets expressed CD40 with CD40 expressed vascular components and may increase inflammatory cell adhesion molecules ICAM-1 and VCAM-1, and white blood cells migration to extravascular space. Independently, platelet adhesion to inflamed intestinal endothelial cells may favor angiogenesis by release of VEGF and platelet-derived growth factor (PDGF), which is also a candidate for therapeutic target for IBD [50]. In the inflamed mucosa of IBD patients de Jong et al [51] presented a decrease of tissue plasminogen activator (t-PA) and increase of urokinase plasminogen activator (u-PA). U-PA, in contrast to t-PA, is less fibrin dependent; thus plasmin generated due to u-PA may act as proinflammatory protease. Cytokines IL-1 and TNF- α are able to cause procoagulant action by induction of tissue factor in ECs, platelets and monocytes; and suppress the anticoagulant potential of thrombomodulin by down-regulation of endothelial protein C receptor, which is sustained normally by ECs [52]. This disruption of the protein C system is also related to elevated adherence of ECs, thus support leukocyte enrolment. Recent investigations has indicated that the protein C pathway is expressed not only in ECs of the intestinal vessels, but also in epithelial cells and plays a significant role in enhancing the integrity of tight junctions [53]. In epithelium of patients with Crohn's disease and UC expression of protein C is changed, which may amplify intestinal permeability [54].

6. Angiogenesis in IBD

Vessel quality are probably critical in pathogenesis of IBD (table II). In fact, newly formed vessels in IBD tissues are strongly disorganized and leaky as indicated by associated edema [55, 56]. The

induction and propagation of angiogenesis during IBD has been partly connected with hypoxia. In fact in IBD mucosa hypoxia-inducible factor-1 and -2 transcriptionally activate the expression of VEGF-A [11]. The VEGF gene may also be turn on by other factors, which are significant in IBD such as growth factor; mainly EGF and TGF β and cytokines e.g. TNF α , IL6 and IL-1 β . [8,57]. We and other were found VEGF higher expression in endothelial cells as well as epithelial cells in IBD- inflamed intestine [5, 58]. Increased VEGF levels in both serum and plasma was observed in patients with IBD which may reflect VEGF overexpression in intestinal inflammatory tissue [56, 58, 59, and 60]. In addition increased VEGF has been found in mucosal extracts obtained from IBD patients as compared to controls [5].

Danese et al [5] first documented the significance of angiogenesis as a novel component both in UC and in Crohn's disease pathogenesis (Fig.1). These authors demonstrated higher density of microvesels as well as VEGF expression within intestinal mucosa of active and inactive UC and Chron's disease phase in comparison with the controls. It seems that VEGF may activate two tyrosine kinase receptors; Flt-1 (fms-like tyrosine kinase-1 receptor) and KDR (kinase domain receptor) [61, 62]. The Flt-1 receptor has closer propinquity to VEGF than KDR receptor. We observed significant increase amount of VEGF gene as well as Flt-1 gene, and higher levels of VEGF and Flt-1 protein in intestinal mucosa of active UC stage in comparison with control group and inactive UC stage, but we found only trace KDR gene expression in active UC phase [58]. Previously German investigators Griga et al. (63 64), have shown higher immunochemical reaction indicated VEGF protein in epithelium, and in lamina propria of colonic tissue in active UC stage than in inactive UC stage and normal colonic tissue, which is in agreement with our results (Fig.2 and Fig.3). In striking contrast Greek investigators Giatromanolaki et al. [11] and Kapsoritakis et al. [65]. Using immunohistochemical methods observed only weak specific reaction for VEGF in endothelial cells and enterocytes in UC inflamed colon. One could postulate that in different geographical regions i.e. northern Europe and Mediterranean area, varied environment and/or unknown genetic pattern may modulate VEGF and its receptors intestinal levels. The endothelial damage may precede epithelial barrier dysfunction in IBD. The investigators at the La University using functional, morphologic, and molecular biologic studies in four animal models of UC indicated that endothelial permeability occurs earlier than epithelial barrier dysfunction that is followed by erosions, ulceration, and inflammation suggesting that endothelial disruption might be critical in disease pathogenesis [66]. In addition Lakatos et al. [67], shown that serum antigen concentration of MMP-9 and tissue inhibitors of metalloproteinases TIMP-1 and TIMP-2 were higher in UC and Chron's disease patients as compared to controls and that MMP-9 and TIMP levels significantly correlated with disease activity. Thus MMPs can contribute to remodeling processes in IBD and serum MMP-9 and TIMP might be used as biomarkers of disease activity.

Elevated levels of other angiogenic growth factors including ba-

sic fibroblast growth factor (bFGF) and platelet-derived growth factor (PDGF) have been found in the inflamed mucosa and in the blood of IBD patients [56, 68, 69, and 70]. Placental growth factor (PIGF), a specific regulator of pathological angiogenesis was found to be increased in the serum of IBD patients [20, 71]. Recently Zhou et al presented proangiogenic effects of PIGF on human intestinal microvascular cells (HIMECs) via PI3K/Akt signaling pathway activation [20].

In relation to angiopoietin system, increased of Ang-2 and Tie-2 was observed in serum of patients with IBD [72]. In later studies it was shown immunologically higher levels of Ang-1 and Ang-2 in epithelia of crypt abscesses from patients with active UC as compared with UC patients with remission [73]. It suggests that angiogenic response is regulated by the Ang/Tie, and this pathway may play a role in the progression of UC. Another unique molecular pattern of ECs from angiogenic vessels represent the expression of adhesion molecules integrins $\alpha v\beta 3$ and $\alpha v\beta 5$ involved with recruitment of T-cells to intestinal mucosa. Importantly higher expression of $\alpha v\beta 3$ has been shown in the mucosa of IBD patients [5, 74].

In addition an effect of kinins in angiogenesis has been suggested. Kinins may mediate angiogenesis by upregulation of basic FGF

via B1 bradykinin receptor and by activation of VEGF synthesis via both B1 and B2 bradykinin receptors [75]. Our data [76] demonstrated B1 and B2 receptors in intestinal tissues of human IBD, and indicated that B1 receptor upregulation is critical for kinin function. Kinins have an ability to stimulate proliferation, and interact with growth factors. Bradykinin as growth factor (also as a member autacoids family) is responsible for capillary permeability and edema. Thus, in all probability kinins may support angiogenesis in IBD although relationship between kinins and cytokines and growth factors in this aspect needs more research.

The chloride channels (ClCs) which are expressed in intestinal mast cells and epithelial cells support tight junction integrity, and regulate chloride ion transport, thus contribute to intestinal barrier function. Recent report indicate that dysregulation of these channels in mast cells may mediate mast cell activation and degranulation and immune cell recruitment in inflamed tissue leading to barrier dysregulation and aggravation some IBD symptoms mainly diarrhea [77]. Importantly the number of mast cells and mast cells tryptase expression (a marker of mast cell degranulation) were found to be increased in colonic mucosa and submucosa in experimental and human IBD [78]. Thus chloride channel modulation in mast cells may have a therapeutic potential in IBD.

Table II: Manifestation of endothelial/vascular responses in IBD.

Endothelial activation and expression of adhesion molecules	leucocyte recruitment platelet adhesion inflammation
VEGF expression	ECs proliferation and migration up-regulation of adhesion molecules immune cells recruitment vascular permeability sprouting of angiogenesis
b FGF expression	sprouting of angiogenesis
PDGF expression	sprouting of angiogenesis vascular coverage
Toll – like receptor expression by EC	regulation of endothelial barrier homeostasis specific receptor for bacterial products
Coagulation activation	platelet adhesion and activation impaired of protein C pathway thrombi formation ischemia
Microvascular dysfunction	granulomatous vasculitis ulceration
Angiogenesis	neovascularization remodeling of vasculature initiation /promotion of inflammation

Abbreviations: ECs; endothelial cells, VEGF; vascular endothelial growth factor, b FGF ; basic fibroblast growth factor, PDGF; platelet-derived growth factor.

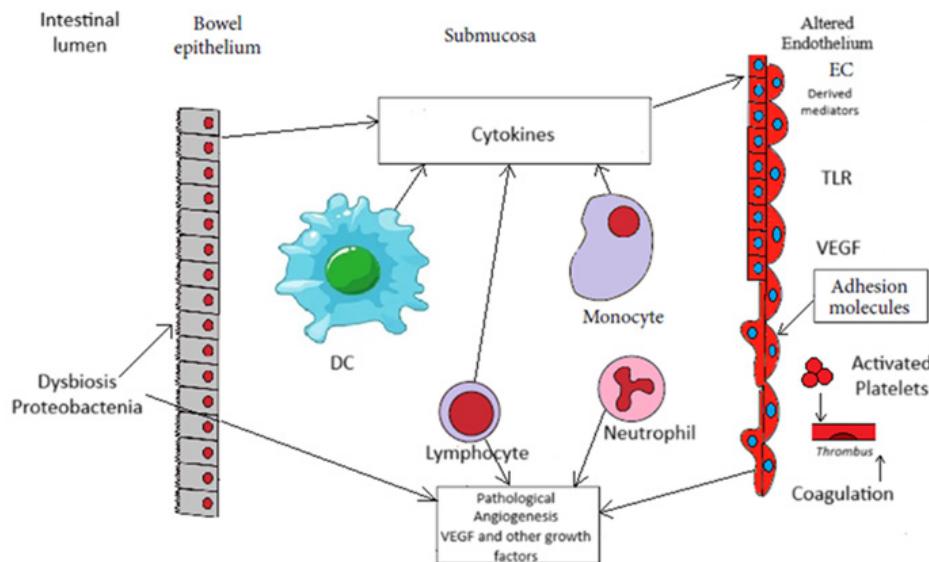


Figure 1: Main factors driving angiogenesis in inflammatory bowel disease. Insight into bowel lumen, intestinal inflamed mucosa/submucosa, and activated and altered endothelium. Infiltrating neutrophils (N), monocytes/macrophages (M), lymphocytes (L), and dendritic cells (DC) secreted cytokines, growth factors, and other factors to stimulate EC (endothelial cells). Activated ECs produced vascular endothelial growth factor (VEGF), expressed of Toll – like receptor (TLR), the specific receptor for bacterial products. Activated ECs with expression of adhesion molecules cause leukocyte recruitment to mucosa/ submucosa and platelet adhesion, whereas activation of coagulation and platelets cause thrombi formation in microvasculature, and in turn ischemia. Vascular dysfunction leads to neovascularization and vascular remodeling which characterize pathological angiogenesis.

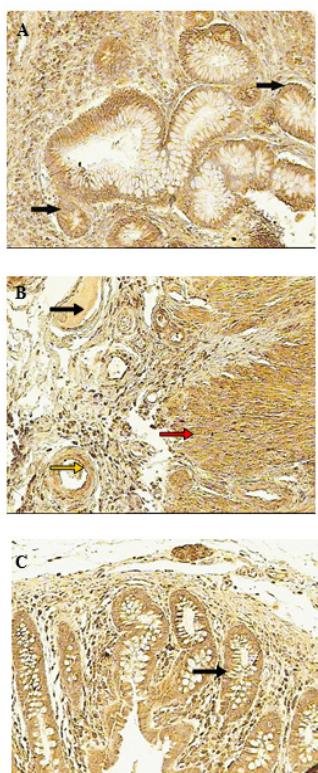


Figure 2: VEGF immunohistochemical localization in UC - inflamed intestine and normal colon (100x). A: Colon in active UC phase, arrows mark specific staining reaction within enterocytes. B: Colon in active UC phase, yellow arrow - specific staining reaction in endothelium of vessel, black arrow - specific staining reaction in lumen vessel, red arrow - specific staining reaction within intestinal smooth muscle cells. C: Normal colon, arrow marks specific staining reaction in enterocytes ((from Reference 58; Fryszt-Naglak D. et al, with permission of Elsevier publishing).

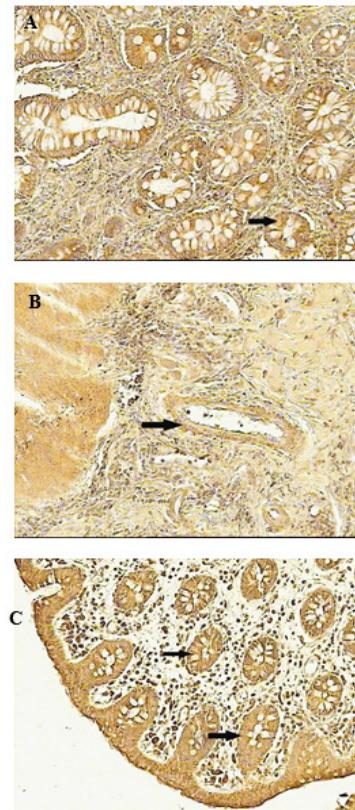


Figure 3: Flt-1 receptor localization in UC - inflamed intestine and normal colon (100x). A: Colon in active UC phase, arrow marks specific staining reaction in enterocytes. B: Colon in active UC phase, arrow marks specific staining reaction in endothelium. C: Normal colon, arrows mark specific staining reaction in enterocytes (from Ref.58; Fryszt-Naglak D. et al, with permission of Elsevier publishing).

6.1. Angiogenesis and Microbiome in IBD

Healthy intestinal bacteria consist mainly of Firmicutes, Bacteroidetes, Actinobacteria, and Proteobacteria which Firmicutes and Bacteroidetes are dominant [79]. Intestinal tissue injury and break of the intestinal barrier lead to the dysregulation of the intestinal milieu and the reactions between the mucosal components and the commensal microbiota. This disequilibrium influence the function of the barrier and it also may alter of the commensal microbiota composition towards dysbiosis [2, 80]. In fact, recent reports have indicated alterations in gut microbiome in IBD towards a less heterogeneous gut microbial composition due to a decrease in commensal anaerobic bacteria; Firmicutes and Bacteroidetes and an increase in the abundance of gram-negative Proteobacteria [81]. Dysbiosis in IBD is characterized by changes in microbiome profile and also by metabolic derangements, mainly decreased short-chain fatty acid (SCFA), and enhanced hydrogen sulfide (H₂S) production, alteration of bile acid and tryptophan metabolism leading to the impairment of epithelial homeostasis and inflammation. Interestingly, relationship of the gut – vascular barrier with the microbiota was shown to increase angiogenesis as observed in human IBD and murine models of colitis [40, 82, 83]. In fact, intestinal microbes are source of angiogenic activators in the form of ligands for Toll-like receptors (TLRs) and protease – activated receptors (PARs) [82, 83]. Ex vivo activated intestinal vessels as well as human intestinal microvascular cells (HIMECs) exposed to microbial products may sprout angiogenesis. These effects are mediated through Toll-like and NOD-like receptors (TLRs and NLRs) which represent innate immune responses [84]. Expression of TLRs on vascular endothelial cells is upregulated by vascular inflammation as well as by bacterial lipopolysaccharide (LPS). Recently it has been shown that activation of TLRs and NLRs by specific bacterial ligands selectively upregulates the levels of carcinoembryonic antigen-related cell adhesion molecule 1 (CEACAM1) produced by HIMECs, and simultaneously induces angiogenesis. These results suggest that a cooperation of endogenous and exogenous innate immune factors by reciprocal action of CEACAM1 and microbiota is essential to promote intestinal angiogenesis in IBD [85]. Microbial biomarkers are emerging as promising non-invasive tools to predict disease activity' disease risk, and recurrence after surgery. The most cost-effective and less invasive method is the analysis of bacterial DNA in plasma. Importantly, bacterial genomic fragments (bactDNA) was observed in the blood for up to 50% of IBD patients [86], and bacterial DNA translocation can be regarded as risk factor of relapse at 6 months in patients with Crohn's disease, and also independently it is connected with an increased risk of hospitalization and steroids induction [87]. Recent report indicate combination of sequencing techniques (methylation-based human cell-specific profiling together with shotgun metagenomics) to characterize the human and the microbial DNA content in feces. Combining neutrophil and other cell type fecal DNA fractions was demonstrated as non-invasive way to distinguish between inactive and active IBD and better disease monitoring [88]. Other approach which may indicate potential targets to aid diagnosis and direct therapeutic options in

IBD refers to stool 16S rRNA gene sequencing and multi-system metabolomic phenotyping (using nuclear magnetic and mass spectroscopy), with subsequent integrative network analysis to delineate novel microbiota -metabolome interactions [89].

In addition some authors have reported endotoxemia and LPS-binding protein in systemic circulation of IBD patients [91, 92]. Of note a link exist between the gut – vascular barrier and the blood–brain barrier (BBB) dysfunction in IBD. Depression and anxiety disorders have been demonstrated in up to 40% of patients with active IBD [93], and less frequently deterioration in cognitive functions [93, 94]. Thus circulating bacterial products and pro-inflammatory mediators in IBD patients may enhance central nervous system disorders. It is documented that in IBD microbiota and their products may modulate the function of immune cells as well as non –immune cells, mainly ECs, thus play a role in initiation or progression of inflammation. Importantly, in IBD microbial products via activating TLRs in ECs may trigger angiogenesis. Thus, it is fascinating to suggest that decrease of the gut microbial load might suppress angiogenesis.

6.2. Treatment of Angiogenesis in IBD

The TNF- α as central mediator to induce the inflammatory reactions has been documented in Crohn's disease and in UC. Thus anti-TNF- α compounds are currently in the biological treatment of moderate and active Crohn's disease and UC [95]. Italian investigators in clinical study have presented the ability of infliximab to affect mucosal angiogenesis in patients with Crohn's disease demonstrated that administration of infliximab downregulates mucosal angiogenesis suggesting that inflammation-driven angiogenesis in the gut mucosa may contributes to the therapeutic efficacy of TNF- α blockade of [96]. Importantly Algaba et al. [97], have shown that circulating VEGF and angiopoietin -1 levels decreased during anti – TNF therapy in UC and Crohn's disease patients [97]. Whether these changes are a signs of improvement is a matter of debate. In addition, the treatment with infliximab downregulates the CD40/CD40L pathway in patients with Crohn's disease [98], which may improve endothelial dysfunction reducing thrombi formation in intestinal vasculature. Therapeutic strategies targeting VEGF represent an interesting way to reduce intestinal angiogenesis and mucosal inflammation. Upregulation of VEGF, a potent vascular permeability factor during early stages of UC may provide evidence to impaired endothelial barrier. Tolstanova et al [99] showing in experimental rat model of UC that anti-VEGF antibody diminished colonic vascular permeability and improved morphologic signs of colitis which indicate the pathogenic role of VEGF. Other pro-angiogenic growth factor e.g. placental growth factor (PIGF) is upregulated in IBD patients via activation of tyrosine kinase -phosphoinositide 3-kinase / protein kinase B (PI3K/ Akt) signaling pathway. Thus PIGF/PI3K/Akt signaling can be as an appropriate therapeutic target in IBD [20]. Bevacizumab a humanized IgG1 monoclonal antibody directed against VEGF-A is used as anti-angiogenic agent in various cancers including colorectal cancer. However up to now there are only case reports showing beneficial effect of bevacizumab (anti-VEGF-antibody)

and sunitinib (multiple tyrosine kinase inhibitor of VEGF and PDGF) treatment in patients with Crohn's disease [100,101]. Importantly, unwanted effects of bevacizumab may contain impaired wound healing, bleeding and even intestinal perforation, although such complications are rare [102,103]. Moreover in the light of case report of Loriot et al. [104] anti – angiogenic treatment may exacerbate IBD. Recently safety of bevacizumab in association with chemotherapy in colonic cancers coexisting with IBD has been retrospectively evaluated. The authors have documented that bevacizumab is safe in cancer patients with IBD, and no clinical IBD exacerbation were found during bevacizumab treatment [105].

Other option for vessel-directed therapy of IBD are anti-adhesion molecules, such as vedolizumab and etrolizumab [106]. Vedolizumab, the anti-integrin- $\alpha 4\beta 7$ -specific antibody may induce long-term remission in refractory patients with Crohn disease and UC [107,108]. This drug blocks reactions between $\alpha 4\beta 7$ and mucosal adhesion molecule (MAdCAM) involved in leukocyte recruitment, whereas etrolizumab, a monoclonal antibody against MAdCAM-1 acts as targets for the $\beta 7$ subunit suppresses accumulation of T lymphocytes [109]. Of note Danese et al. in an experimental mouse model of ulcerative colitis have been shown inhibition angiogenesis employing the integrin $\alpha v\beta 3$ inhibitor (ATN161) and at the same time decrease of intestinal inflammatory alterations [110]. However it should be emphasize that a beneficial effect of anti-angiogenic drugs as additional IBD treatment are still controversial. In fact up to now no clinical trial has been performed using anti anti-angiogenic treatment in IBD patients.

Among possible microbiota-targeted interventions, pro-biotics application provide limited effectiveness, in part because currently the limited microbial species available as probiotics. In contrast, fecal microbiota transplantation (FMT) replace intestinal milieu of a recipient by fecal solution from a donor [111]. Successful results from the use of FMT for recurrent *Clostridium difficile* infection indicated new approaches for IBD to replace existing microbiota and enhance protective bacterial numbers and metabolism. A number of clinical trials in patients with UC and Crohn's disease showed the safety but variable efficacy of FMT in achieving clinical remission and maintaining remission after being achieved via drug therapy [112]. Involvement of angiogenesis in efficacy of FMT was indicated in murine irradiation model. In this model was found that application of FMT alleviated and protected against radiation induced intestinal injury via upregulation of intestinal VEGF expression [113]. Expanding insights into donor microbial composition, recipient factors, and post-transplant microbial profiles may define frequency of FMT administration and predict successful FMT outcomes.

6.3. Dual role of Angiogenesis

The data of many studies indicate the significance of angiogenesis in IBD which may initiate and/or amplify chronic inflammation and fibrosis in the intestine. VEGF induces an abnormal angiogenesis, and other growth factors mainly placental growth factor (PIGF), basic epidermal growth factor (bFGF) and transforming growth factor- β (TGF- β) may play a critical role in pathogenesis

of IBD. On the other hand these growth factors can be favorable in healing and reparation processes [114]. It has been shown in experimental studies that administration of PDGF or FGF significantly accelerated healing of UC [115]. In fact EGF enemas have been demonstrated to be useful in patients with UC [116]. The researchers at the La University documented in rats models that the molecular mechanisms of bFGF in UC healing is connected with increased cell proliferation, especially angiogenesis and simultaneously decrease of cytokines and inflammatory cells [117]. Other authors also demonstrated that activation of Rac1 (a key factor of the Rho guanosine triphosphatase (GTPase) family) improved VEGF-induced angiogenesis in vivo as indicated by measurement of vascular density and diminished vessel leakiness in an angiogenic model [118]. Moreover anti-angiogenic factors are upregulated in inflamed intestine such as thrombospondin, endostatin-a cleaved fragment of collagen XVIII and angiostatin—a cleaved fragment of plasminogen [28,119]. Thus, therapeutic inhibition of angiogenesis may reduce wound healing in experimental colitis models and IBD.

References

1. Guan Q A. Comprehensive Review and Update on the Pathogenesis of Inflammatory Bowel Disease. *J Immunol Res.* 2019; 7247238.
2. Ni J, Wu GD, Albenberg L, Tomov VT. Gut microbiota and IBD: Causation or correlation? *Nat Rev Gastroenterol Hepatol.* 2017; 14: 573-584.
3. Benelli R, Lorusso G, Albini A, Noonan DM. Cytokines and chemokines as regulators of angiogenesis in health and disease. *Current Pharmaceutical Design.* 2006; 12: 3101-3115.
4. Sajib S, Zahra FT, Lionakis MS. Mechanisms of angiogenesis in microbe-regulated inflammatory and neoplastic conditions. *Angiogenesis.* 2018; 21: 1-14.
5. Danese S, Sans M, Motte C. Angiogenesis as a novel component of inflammatory bowel disease pathogenesis. *Gastroenterology.* 2006; 130: 2060-2073.
6. Ferrara N, Gerber HP. The biology of VEGF and its receptors. *Nature Medicine.* 2003; 9: 669-676.
7. Cebe-Suarez S, Zehnder-Fjällman, A.; Ballmer-Hofer, K. The role of VEGF receptors in angiogenesis; complex partnerships. *Cell Mol Life Sci.* 2006; 63: 601-15.
8. Ferrara, N. Vascular endothelial growth factor: basic science and clinical progress. *Endocr Rev.* 2004; 25: 581-611.
9. Felmeden DC, Blann AD. Angiogenesis: basic patho-physiology and implications for disease. *Eur Heart J.* 2003; 24: 586-603.
10. Dachs GU, Tozer GM. Hypoxia modulated gene expression:angiogenesis, metastasis and therapeutic exploitation. *Eur J Cancer.* 2000; 36: 1649-60.
11. Giatromanolaki A, Sivridis E. Hypoxia inducible factor 1alpha and 2alpha overexpression in inflammatory bowel disease. *J Clin Pathol.* 2003; 56: 209-213.
12. Paleolog EM, Young S, Stark AC. Modulation of angiogenic vascular endothelial growth factor by tumor necrosis factor alpha and interleukin-1 in rheumatoid arthritis. *Arthritis Rheum.* 1998; 41: 1258-1265.

13. Bottomley MJ, Webb N J. Peripheral blood mononuclear cells from patients with rheumatoid arthritis spontaneously secrete vascular endothelial growth factor (VEGF): specific up-regulation by tumour necrosis factor-alpha (TNF-alpha) in synovial fluid. *Clin Exp Immunol.* 1999; 117: 171-176.
14. Biddlestone J, Bandarra D, Rocha S. The role of hypoxia in inflammatory disease. *Inter J Mol Med.* 2015; 35: 859-869.
15. Bakirtzi K, West G, Fiocchi C. The neurotensin-HIF-1 α -VEGF α axis orchestrates hypoxia, colonic inflammation, and intestinal angiogenesis. *Amer J Pathol.* 2014; 184: 3405-3414.
16. Alvarez RH, Kantarjian HM. Biology of platelet-derived growth factor and its involvement in disease. *Mayo Clinic Proc.* 2006; 81: 1241-1257.
17. Ierardi E, Giorgio F, Zotti M. Infliximab therapy downregulation of basic fibroblast growth factor/syndecan 1 link: a possible molecular pathway of mucosal healing in ulcerative colitis. *J Clin Pathol.* 2011; 64: 968-972.
18. Kanazawa S, Tsunoda T, Onuma E, Majima T. VEGF, basic-FGF, and TGF- β in Crohn's disease and ulcerative colitis: a novel mechanism of chronic intestinal inflammation. *Amer J Gastroenterol.* 2001; 96: 822-828.
19. Itoh H, Kataoka H. Roles of hepatocyte growth factor activator (HGFA) and its inhibitor HAI-1 in the regeneration of injured gastrointestinal mucosa. *J Gastroenterol.* 2002; 37: 15-21.
20. Zhou Yi, Yuan Zhao Y. Placental growth factor enhances angiogenesis in human intestinal microvascular endothelial cells via PI3K/Akt pathway: Potential implications of inflammation bowel disease. *Biochem Biophys Res Commun.* 2016; 470: 967-74.
21. Carmeliet P. Angiogenesis in life, disease and medicine. *Nature.* 2005; 438: 932-6.
22. Yancopoulos GD, Davis S, Gale NW. Vascular-specific growth factors and blood vessel formation. *Nature.* 2000; 407: 242-8.
23. Kim I, Kim JH. Angiopoietin-2 at high concentration can enhance endothelial cell survival through the phosphatidylinositol 3'-kinase/Akt signal transduction pathway. *Oncogene.* 2000; 19: 4549-52.
24. Lobov IB, Brooks PC. Angiopoietin-2 displays VEGF-dependent modulation of capillary structure and endothelial cell survival in vivo. *Proc Natl Acad Sci USA.* 2002; 99: 11205-10.
25. Fiedler U, Reiss Y, Scharpfenecker M. Angiopoietin-2 sensitizes endothelial cells to TNF- α and has a crucial role in the induction of inflammation. *Nature Medicine.* 2006; 12: 235-239.
26. Linares PM, Chaparro M, Gisbert J. Angiopoietins in inflammation and their implication in the development of inflammatory bowel disease. *J Crohn's and Colitis.* 2014; 8: 183-190.
27. Zak S, Treven J. Lack of thrombospondin-1 increases angiogenesis in a model of chronic inflammatory bowel disease. *Inter J Colorectal Dis.* 2008; 23: 297-304.
28. Danese S. Negative regulators of angiogenesis in inflammatory bowel disease; thrombospondin in the spotlight. *Pathobiology.* 2008; 75: 22-24.
29. Ortiz-Masià D, Díez I. Induction of CD36 and thrombospondin-1 in macrophages by hypoxia-inducible factor 1 and its relevance in the inflammatory process. *PLoS ONE.* 2012; 7.
30. Farhadi A, Banan A, Fields J. Intestinal barrier: an interface between health and disease. *J Gastroenterol Hepatol.* 2003; 18: 479-487.
31. Spadoni I, Zagato E, Bertocchi A. A gut-vascular barrier controls the systemic dissemination of bacteria. *Science.* 2015; 350: 830-834.
32. Brescia P, Rescigno M. The gut vascular barrier: A new player in the gut-liver-brain axis. *Trends Mol. Med.* 2021; 27: 844-855.
33. Mankertz J, Schulzke JD. Altered permeability in inflammatory bowel disease: pathophysiology and clinical implications. *Curr Opin Gastroenterol.* 2007; 23: 379-383.
34. Stürzl M, Kunz M, Krug SM. Angiocrine regulation of epithelial barrier integrity in inflammatory bowel disease. *Front Med (Lausanne).* 2021; 8: 643607.
35. Neurath MF. Cytokines in inflammatory bowel disease. *Nat Rev Immunol.* 2014; 14: 329.
36. Pober JS, Sessa WC. Evolving functions of endothelial cells in inflammation. *Nat Rev Immunol.* 2007; 7: 803-15.
37. Himadri R, Shalini B, Seppo YH. Biology of vascular endothelial growth factors. *FEBS Letters.* 2006; 580: 2879-87.
38. Spadoni I, Fornasa G. Organ-specific protection mediated by cooperation between vascular and epithelial barriers. *Nat Rev Immunol.* 2017; 17: 761-73.
39. Lopez-Posadas R, Stürzl M. Interplay of GTPases and cytoskeleton in cellular barrier defects during gut inflammation. *Front Immunol.* 2017; 8: 1240.
40. Kabouridis PS, Lasrado R, McCallum S, Chng SH. Microbiota controls the homeostasis of glial cells in the gut lamina propria. *Neuron.* 2015; 85: 289-295.
41. Morita K, Sasaki H, Furuse M, Tsukita S. Endothelial claudin: claudin-5/TMVCF constitutes tight junction strands in endothelial cells. *J Cell Biol.* 1999; 147: 185-94.
42. Günzel D, Yu ASL. Claudins and the modulation of tight junction permeability. *Physiol Rev.* 2013; 93: 525-569.
43. Wolburg H, Wolburg-Buchholz K. Localization of claudin-3 in tight junctions of the blood-brain barrier is selectively lost during experimental autoimmune encephalomyelitis and human glioblastoma multiforme. *Acta Neuropathol.* 2003; 105: 586-92.
44. Wakefield A, Sawyer A. Pathogenesis of Crohn's disease: multifocal gastrointestinal infarction. *Lancet.* 1989; 2: 1057-62.
45. Wakefield A, Sabkey EA, Dhillon AP. Granulomatous vasculitis in Crohn's disease. *Gastroenterology.* 1991; 100: 1279-87.46.
46. Yoshida H, Granger DN. Inflammatory Bowel Diseases: a paradigm for the link between coagulation and inflammation. *Inflamm Bowel Dis.* 2009; 15: 1245-55.
47. Dhillon AP, Anthony A. Mucosal capillary thrombi in rectal biopsies. *Histopathology.* 1992; 21: 127-33.
48. Danese S, Katz JA, Saibeni S. Activated platelets are the source of elevated levels of soluble CD40 ligand in the circulation of inflammatory bowel disease patients. *Gut.* 2003; 52: 1435-41.
49. Danese S, de La Motte. Platelets trigger a CD40 – dependent inflammatory response in the microvasculature of inflammatory bowel disease patients. *Gastroenterology.* 2003; 124: 1249-64.
50. Rutela S, Vetrano S, Correale C. Enhanced platelet adhesion induces angiogenesis in intestinal inflammation and inflammatory bowel disease microvasculature. *J Cell Mol Med.* 2011; 15: 625-34.

51. de Jong, Porte RJ. Disturbed fibrinolysis in patients with inflammatory bowel disease. A study of blood plasma, colon mucosa, and faeces. *Gut*. 1989; 30: 188-94.
52. Van de Wouwer M, Collen D, Conway EM. Thrombomodulin-protein C-EPCR system: integrated to regulate coagulation and inflammation. *Arterioscler Thromb Vasc Biol*. 2004; 24: 1374-1383.
53. Vetrano S, Ploplis VA, Sala E. Unexpected role of anticoagulant protein C in controlling epithelial barrier integrity and intestinal inflammation. *Proc Natl Acad Sci USA*. 2011; 108: 19830-19835.
54. Faioni, EM, Ferrero S. Expression of endothelial protein C receptor and thrombomodulin in the intestinal tissue of patients with inflammatory bowel disease. *Crit Care Med*. 2004; 32: S266-S270.
55. Cromer WE, Mathis JM, Granger DN. Role of the endothelium in inflammatory bowel diseases. *World J Gastroenterol*. 2011; 17: 578-93.
56. Alkim C, Alkim H. Angiogenesis in inflammatory bowel disease. *Int J Inflam*. 2015; 2015: 970890.
57. Stadnicki A, Machnik G. Transforming growth factor-beta 1 and its receptors in patients with ulcerative colitis. *Int Immunopharmacol*. 2009; 9: 761-69.
58. Frysz-Naglak D, Fryc B, Klimacka-Nawrot E. Expression, localization and systemic concentration of vascular endothelial growth factor (VEGF) and its receptors in patients with ulcerative colitis. *Int Immunopharmacol*. 2011; 11: 220-225.
59. Scaldaferri F, Vetrano S, Sans M. VEGF-A links angiogenesis and inflammation in inflammatory bowel disease pathogenesis. *Gastroenterology*. 2009; 136: 585-595.e5.
60. Bousvaros A, Leichtner A, Zurakowski D. Elevated serum vascular endothelial growth factor in children and young adults with Crohn's disease. *Dig Dis Sci*. 1999; 44: 424-430.
61. Himadri R, Shalini B. Biology of vascular endothelial growth factors. *FEBS Letters*. 2006; 580: 2879-87.
62. Proczka RM, Polański JA. The significance of vascular endothelial growth factor in the neoangiogenesis process. The role of hypoxia in the endothelial cells proliferation process and in the formation of collateral circulation. *Acta Angiol*. 2003; 9: 143-9.
63. Griga T, Gutzeit A, Sommerkamp C. Increased production of vascular endothelial growth factor by peripheral blood mononuclear cells in patients with inflammatory bowel disease. *Eur J Gastroenterol Hepatol*. 1999; 11: 175-179.
64. Griga T, May B, Pfisterer O, Müller KM. Immunohistochemical localization of vascular endothelial growth factor in colonic mucosa of patients with inflammatory bowel disease. *Hepato-Gastroenterology*. 2002; 49: 116-23.
65. Kapsoritakis A, Sfiridakis A. Vascular endothelial growth factor in inflammatory bowel disease. *Int J Colorectal Dis*. 2003; 18: 418-22.
66. Tolstanova G, Deng, X, French SW. Early endothelial damage and increased colonic vascular permeability in the development of experimental ulcerative colitis in rats and mice. *Lab Invest*. 2012; 92: 9-21.
67. Lakatos G, Hritz I, Varga MZ. The impact of matrix metalloproteinases and their tissue inhibitors in inflammatory bowel diseases. *Dig Dis Sci*. 2012; 57: 289-295.
68. Krzystek-Korpacka M, Neubauer K. Platelet-derived growth factor-B reflects clinical, inflammatory and angiogenic disease activity and oxidative stress in inflammatory bowel disease. *Clin Biochem*. 2009; 42: 1602-1609.
69. Macé V, Ahluwalia A, Coron E. Confocal laser endomicroscopy: A new gold standard for the assessment of mucosal healing in ulcerative colitis. *J Gastroenterol Hepatol*. 2015; 30: 85-92.
70. Ippolito C, Colucci R. Fibrotic and Vascular Remodelling of Colonic Wall in Patients with Active Ulcerative Colitis. *J Crohn's Colitis*. 2016; 10: 1194-1204.
71. Pousa ID, Maté J, Salcedo-Mora X. Role of vascular endothelial growth factor and angiopoietin systems in serum of Crohn's disease patients. *Inflamm Bowel Dis*. 2008; 14: 61-67.
72. Kourtroubakis IE, Xidakis C, Karmiris K. Potential role of soluble angiopoietin-2 and Tie-2 in patients with inflammatory bowel disease. *Eur J Clin Invest*. 2006; 36: 127-32.
73. Yoshizaki A, Nakayama T, Naito S. Expression patterns of angiopoietin-1, -2, and tie-2 receptor in ulcerative colitis support involvement of the angiopoietin/tie pathway in the progression of ulcerative colitis. *Dig Dis Sci*. 2009; 54: 2094-9.
74. Dotan I, Allez M. The role of integrins in the pathogenesis of inflammatory bowel disease: Approved and investigational anti-integrin therapies. *Med Res Rev*. 2020; 40: 245-262.
75. Colman RW. Regulation of angiogenesis by the kallikrein-kinin system. *Cur Pharm Des*. 2006; 12: 599-607.
76. Stadnicki A. Intestinal tissue kallikrein – kinin system in inflammatory bowel disease. *Inflamm Bowel Dis*. 2011; 17: 645-654.
77. Aljameeli AM, Aswaid B, Bharati B, Gohri V. Chloride channels and mast cell function: pioneering new frontiers in IBD therapy. *Mol Cell Biochem*. 2025; 480: 3951-3969.
78. Shao-Heng He. Key role of mast cells and their major secretory products in inflammatory bowel disease. *World J Gastroenterol*. 2004; 10: 309-318.
79. Khandagale A, Reinhardt C. Gut microbiota - architects of small intestinal capillaries. *Front Biosci*. 2018; 23: 752-766.
80. Mishima Y, Sartor RB. Manipulating resident microbiota to enhance regulatory immune function to treat inflammatory bowel diseases. *J Gastroenterol*. 2020; 55: 4-14.
81. Alam MT, Amos GCA. Microbial imbalance in inflammatory bowel disease patients at different taxonomic levels. *Gut Pathol*. 2020; 12:1.
82. Reinhardt C, Bergentall M, Greiner, TU. Tissue factor and PAR1 promote microbiota-induced intestinal vascular remodelling. *Nature*. 2012; 483: 627-631.
83. Schirbel A, Kessler S, Rieder F. Pro-angiogenic activity of TLRs and NLRs: A novel link between gut microbiota and intestinal angiogenesis. *Gastroenterology*. 2013; 144: 613-623.
84. Shin HS, Xu F, Bagchi A. Bacterial lipoprotein TLR2 agonists broadly modulate endothelial function and coagulation pathways in vitro and in vivo. *J Immuno*. 2011; 186: 1119-1130.
85. Schirbel A, Rebert N. Mutual Regulation of TLR/NLR and CEA-CAM1 in the Intestinal Microvasculature: Implications for IBD Pathogenesis and Therapy. *Inflamm Bowel Dis*. 2019; 25: 294-305.

86. Gutiérrez A, Francés R. Cytokine association with bacterial DNA in serum of patients with inflammatory bowel disease. *Inflamm. Bowel Dis.* 2009; 15: 508-514.
87. Gutiérrez A, Zapater P, Juanola O. Gut Bacterial DNA Translocation is an Independent Risk Factor of Flare at Short Term in Patients with Crohn's Disease. *Am J Gastroenterol.* 2016; 111: 529-540.
88. Mazzoni Ch, Ochana B, Focht G. Human DNA levels in feces reflect gut inflammation and associate with presence of gut species in IBD patients across the age spectrum. *Res Sq.* 2025; 6809327.
89. Radhakrishnan ST, Mullish BH, Olbei ML. Deciphering the microbiome-metabolome landscape of an inflammatory bowel disease inception cohort. *Gut Microbes.* 2025; 2527863.
90. Gardiner KR, Halliday MI, Barclay GR. Significance of systemic endotoxemia in inflammatory bowel disease. *Gut.* 1995; 36: 897- 901.
91. Rojo ÓP, Román ALS, Arbizu EA. Serum lipopolysaccharide-binding protein in endotoxemic patients with inflammatory bowel disease. *Inflamm Bowel Dis.* 2006; 13: 269-277
92. Navabi S, Gorrepati VS, Yadav S, Chintanaboina J. Influences and Impact of Anxiety and Depression in the Setting of Inflammatory Bowel Disease. *Inflamm Bowel Dis.* 2018; 24: 2303-2308.
93. Byrne G, Rosenfeld G, Leung Y. Prevalence of Anxiety and Depression in Patients with Inflammatory Bowel Disease. *Can J Gastroenterol Hepatol.* 2017; 6496727.
94. Tadin Hadjina I, Zivkovic PM, Matetic A. Impaired neurocognitive and psychomotor performance in patients with inflammatory bowel disease. *Sci Rep.* 2019; 9: 13740.
95. Błoński W, Stadnicki A, Lichtenstein G, Burke A. Infliximab use in ulcerative colitis. In: *Ulcerative Colitis. The complete quid to medical management.* Ed: GR Lichtenstein, EJ Scherl New York Slack Inc. 2011; 237-254.
96. Rutella S, Fiorino G, Vetrano S. Infliximab therapy inhibits inflammation-induced angiogenesis in the mucosa of patients with Crohn's disease. *Am J Gastroenterol.* 2011; 106: 762-70.
97. Algaba A, Linares PM, Fernández-Contreras M, Figuerola A.. The effects of infliximab or adalimumab on vascular endothelial growth factor and angiopoietin 1 angiogenic factor levels in inflammatory bowel disease: serial observations in 37 patients. *Inflamm Bowel Dis.* 2014; 20: 695-702.
98. Danese S, Sans M. TNF - α blockade down – regulates the CD40/ CD40L pathway in the mucosal microcirculation : a novel anti – inflammatory mechanism of infliximab in Crohn's disease. *J Immunol.* 2006; 176: 2617-2624.
99. Tolstanova G, Khomenko T, Deng X. Neutralizing anti-vascular endothelial growth factor (VEGF) antibody reduces severity of experimental ulcerative colitis in rats: direct evidence for the pathogenic role of VEGF. *J Pharmacol Exp Ther.* 2009; 328: 749-757.
100. Magro F, Costa C. Long-standing remission of Crohn's disease under imatinib therapy in a patient with Crohn's disease. *Inflamm Bowel Dis.* 2006; 12: 1087-89.
101. Coriat R, Mir O, Leblanc S. Feasibility of anti-VEGF agent bevacizumab in patients with Crohn's disease. *Inflamm Bowel Dis.* 2010; 17: 1632.
102. Hapani S, Chu D, Wu S. Risk of gastrointestinal perforation in patients with cancer treated with bevacizumab: A meta-analysis. *Lancet Oncol.* 2009; 10: 559-568.
103. Burger RA, Brady MF, Bookman MA. Risk Factors for GI Adverse Events in a Phase III Randomized Trial of Bevacizumab in First-Line Therapy of Advanced Ovarian Cancer: A Gynecologic Oncology Group Study. *J Clin Oncol.* 2014; 32: 1210-1217.
104. Loriot Y, Boudou-Rouquette P. Acute exacerbation of hemorrhagic rectocolitis during antiangiogenic therapy with sunitinib and sorafenib. *Ann Oncol.* 2008; 19: 1975.
105. Herrera-Gómez RG, Grecea M, Gallois C. Safety and Efficacy of Bevacizumab in Cancer Patients with Inflammatory Bowel Disease. *Cancers (Basel).* 2022; 14: 2914.
106. Löwenberg, M.; D'Haens, G. Next-Generation Therapeutics for IBD. *Curr Gastroenterol Rep.* 2015; 17: 21.
107. Feagan BG, Rutgeerts P, Sands BE. Vedolizumab as induction and maintenance therapy for ulcerative colitis. *N Engl J Med.* 2013; 369: 699-710.
108. Sandborn WJ, Feagan BG. Vedolizumab as induction and maintenance therapy for Crohn's disease. *N Engl J Med.* 2013; 369: 711-721.
109. Zundler S, Schillinger D, Fischer A. Blockade of alpha Ebeta7 integrin suppresses accumulation of CD8+ and Th9 lymphocytes from patients with IBD in the inflamed gut *in vivo.* *Gut.* 2017; 66: 1936-1948.
110. Danese S, Sans M, Spencer D. Angiogenesis blockade as a new therapeutic approach to experimental colitis. *Gut.* 2007; 56: 855-62.
111. Biazzo M, Deidda D. Fecal microbiota transplantation as new therapeutic avenue for human diseases. *J Clin Med.* 2022; 11: 4119.
112. Sartor B. Beyond random fecal microbial transplants: next generation personalized approaches to normalize dysbiotic microbiota for treating IBD. *Gastrenterol Clin North Amer.* 2025; 54: 333-350.
113. Cui M, Xiao H, Li Y, Zhou. Faecal microbiota transplantation protects against radiation-induced toxicity. *EMBO Mol Med.* 2017; 9: 448-461.
114. Dignass AU, Podolsky DK. Epithelial restitution and intestinal repair. In: Sartor RB, Sandborn WJ, editors. *Kirsner's inflammatory bowel diseases.* Philadelphia: Saunders. 2004; 18-29.
115. Deng X, Szabo S, Khomenko Y. Novel pharmacologic approaches to the prevention and treatment of ulcerative colitis. *Curr Pharm Des.* 2013; 19: 7-28.
116. Sinha A, Nightingale J, West KP. Epidermal growth factor enemas with oral mesalamine for mild-to-moderate left – sides ulcerative colitis or proctitis. *N Engl J Med.* 2003; 349: 350-57.
117. Paunovic B, Deng X, Khomenko T. Molecular Mechanisms of Basic Fibroblast Growth Factor Effect on Healing of Ulcerative Colitis in Rats. *J Pharma Exp Ther.* 2011; 339: 430-437.
118. Hoang MV, Nagy JA. Active Rac1 improves pathologic VEGF neovessel architecture and reduces vascular leak: mechanistic similarities with angiopoietin-1. *Blood.* 2011; 117: 1751-60.
119. Singh UP, Singh NP, Murphy EA. Chemokine and cytokine levels in inflammatory bowel disease patients. *Cytokine.* 2016; 77: 44-49.